

SEPARATION OF GINKGOLIDES AND BILOBALIDE FROM G. BILOBA

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This application claims priority of U.S. Provisional Application No. 60/519,840 filed November 12, 2003, the contents of which are hereby incorporated by reference into this application.

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This invention has been made with government support under National Institutes of Health grant AI10187. Accordingly, the U.S. Government has certain rights in the invention.

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Throughout this application various publications are referenced in parenthesis. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

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Background of the Invention

The *Ginkgo biloba* tree mentioned in the Chinese *Materia Medica* 5000 years ago (Deng, Q. *Drug Use Perspective* 1988, 1, 57-58) has gained wide interest for its biological activities, especially in the treatment of memory related afflictions. Standardized extract EGB761 (Drieu, K.; Jaggy, H. in *Ginkgo biloba*; van Beek, T.A., Ed.; Harwood Academic Publishers: Amsterdam, 2000; Vol. 12, pp 267-277) from *G. biloba* leaves is a complex mixture containing flavonoids (22 - 24%) and terpene trilactones (TTL, 5 - 7%), and is one of the best selling herbal medicines (Schmid, W. *Nature (London)* 1997, 386, 755). The structures of ginkgolides A, B, and C were determined in 1967 (Figure 1), (Nakanishi, K. *Pure Appl. Chem.* 1967, 14, 89-113; Okabe, K.; Yamada, K.; Yamamura, S.; Takada, S. *J. Chem.*

Soc. C 1967, 2201-2206) while those of ginkgolide J (Weinges, K.; Hepp, M.; Jaggy, H. Liebigs Ann. Chem. 1987, 521-526) and bilobalide (Nakanishi, K.; Habaguchi, K.; Nakadaira, Y.; Woods, M.C.; Maruyama, M.; Major, R.T.; Alauddin, M.; Patel, A.R.; Weinges, K.; Bahr, W. J. Am. Chem. Soc. 1971, 93, 3544-3546) were elucidated later. Flavonoids and TTLs are believed to be associated with most of the pharmacological properties of *G. biloba* extracts. While flavonoids can be obtained from many other plants, ginkgolides and bilobalide are unique components of *G. biloba* extracts. They have attracted scientists to investigate the biological activity and putative link of these compounds to pharmacological activities of EGb761, particularly those related to neuromodulation (long term potentiation, cognitive symptoms of dementia) (MacLennan, K.M.; Darlington, C.L.; Smith, P.F. *Progress in Neurobiology* (Oxford, United Kingdom) 2002, 67, 235-257). Ginkgolides are well known for their platelet activating factor antagonistic activity (Braquet, P. *Drugs of the Future* 1987, 12, 643-699) and recently also for the selective inhibition of glycine receptors (Kondratskaya, E.L.; Krishtal, O.A. *Neurophysiology* (Translation of *Neirofiziologiya*) 2002, 34, 155-157). Although bilobalide lacks these activities its neuroprotective properties have been reported (Zhou, L.J.; Zhu, X.Z. *J. Pharmacol. Exp. Ther.* 2000, 293, 982-988; Janssens, D.; Remacle, J.; Drieu, K.; Michiels, C. *Biochem. Pharmacol.* 1999, 58, 109-119; Chandrasekaran, K.; Mahrabian, Z.; Spinnewyn, B.; Drieu, K.; Fiskum, G. *Brain Research* 2001, 922, 282-292). Recently, methods for the extraction of TTLs with organic solvents (PCT International Application No. WO 02/083158, published October 24, 2002 (Teng, B.P.)), water (Lichtblau, D.; Berger, J.M.; Nakanishi, K. *Journal of Natural Products* 2002, 65, 1501-1504; U.S. Patent No. 6,590,109, issued July 8, 2003 to Lichtblau, et al.)), pressurized water (U.S. Patent No. 6,524,628, issued February 25, 2003 to Wai, et al.) or

supercritical fluids (Choi, Y.H.; Kim, J.; Yoo, K.P. *Chromatographia* **2002**, *56*, 753-757; van Beek, T.A.; Taylor, L.T. *Phytochemical Analysis* **1996**, *7*, 185-191) have been developed. The individual terpene components can be separated from these enriched extracts by a 10-15 step fractional recrystallization(Nakanishi, K. *Pure Appl. Chem.* **1967**, *14*, 89-113; Okabe, K.; Yamada, K.; Yamamura, S.; Takada, S. *J. Chem. Soc. C* **1967**, 2201-2206; Zhao, J.; Muhammad, I.; Dunbar, D.C.; Khan, I.A.; Fischer, N.H.; Fronczek, F.R. *Acta Crystallographica, Section C.: Crystal Structure Communications* **2002**, C58, o195-o198; Dupont, L.; Dideberg, O.; Germain, G.; Braquet, P. *Acta Cryst.* **1986**, C42, 1759-1762; Sbit, M.; Dupont, L.; Dideberg, O.; Braquet, P. *Acta Cryst.* **1987**, C43, 2377-2381) repeated column chromatography(Weinges, K.; Bahr, W. *Liebigs Ann. Chem.* **1972**, *759*, 158-172), reversed-phase HPLC(Wada, K.; Sasaki, K.; Miura, K.; Yagi, M.; Kubota, Y.; Matsumoto, T.; Haga, M. *Biological and Pharmaceutical Bulletin* **1993**, *16*, 210-212; Lobstein-Guth, A.; Briancon-Scheid, F.; Anton, R. *Journal of Chromatography* **1983**, *267*, 431-438; Camponovo, F.F.; Wolfender, J.L.; Maillard, M.P.; Potterat, O.; Hostettmann, K. *Phytochemical Analysis* **1995**, *6*, 141-148), chromatography system with sephadex LH-20 (Teng, B.P. In *Ginkgolides - Chemistry, Biology, Pharmacology*; Braquet, P., Ed.; J.R. Prous Science Publishers, S.A., **1988**, pp 37-42) or less problematically by chromatography on NaOAc impregnated silica gel(van Beek, T.A.; Lelyveld, G.P. *J. Nat. Prod.* **1997**, *60*, 735-738).

In contrast to the large number of methods in which the separation procedure is rather complicated or the individual terpene trilactones can only be obtained in low purity and/or low quality, described herein are simple and efficient separation methods for the separation of five terpene trilactones from *G. biloba* extract by a combination of

benzylation, chromatography, and hydrogenolysis or a combination of extraction, chromatography and crystallization. Of particular importance for large-scale isolation of ginkgolides as phytopharmaceuticals, the described method 5 enables separation without resorting to preparative reversed-phase HPLC or other expensive procedures.

Summary of the Invention

The subject invention provides a process for separating a terpene trilactone from Ginkgo biloba plant material or from
5 an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

- a) subjecting the Ginkgo biloba plant material or the extract to column chromatography with an appropriate solvent system to produce at least a first fraction containing the terpene trilactone bilobalide, a second fraction eluted after the first fraction containing the terpene trilactones GA and GB, and a third fraction eluted after the second fraction containing at least a preponderance of the terpene trilactones GC and GJ; and
- b) alkylating the terpene trilactone GB of the second fraction so as to produce a first mixture including terpene trilactone GA and alkylated terpene trilactone GB; or alkylating the terpene trilactone GC of the third fraction so as produce a second mixture including terpene trilactone GJ and alkylated terpene trilactone GC, so as to thereby isolate a terpene trilactone.

The invention also provides a process for separating a mixture of terpene trilactones GA and GB comprising the steps of:

- a) dissolving the mixture in a suitable solvent;
- b) adding benzyl bromide and a suitable base to the mixture, thereby preferentially benzylating terpene trilactone GB;

- c) separating benzylated terpene trilactone GB from terpene trilactone GA by column chromatography; and
- 5 d) deprotecting benzylated terpene trilactone GB by catalytic hydrogenation.

The invention further provides a process for separating a mixture of terpene trilactones GC and GJ comprising the steps
10 of:

- a) dissolving the mixture in a suitable solvent;
- b) adding benzyl bromide and a suitable base to the mixture, thereby preferentially benzylating terpene trilactone GC;
- 15 c) separating benzylated terpene trilactone GC from terpene trilactone GJ by column chromatography; and
- d) deprotecting benzylated terpene trilactone GC by catalytic hydrogenation.

The invention also provides a process for separating a terpene trilactone from Ginkgo biloba plant material or an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

- a) subjecting the Ginkgo biloba plant material or the extract to column chromatography with hexane/ethyl acetate to produce at least a first fraction containing the terpene trilactone bilobalide, a second fraction eluted after the first fraction containing the terpene trilactone GA and GB, and a

third fraction eluted after the second fraction containing at least a preponderance of the terpene trilactones GC and GJ;

5 b) rinsing the first fraction of step a) with ethyl ether to produce the terpene trilactone bilobalide in at least 98% purity;

10 c) removing solvent under vacuum from the second fraction in step a) so as to produce a first residue containing at least a preponderance of terpene trilactones GA and GB; admixing the first residue in DMF with K₂CO₃ and benzyl bromide, thereby producing a first reaction mixture; quenching the first reaction mixture with HCl, thereby producing a first quenched product; extracting the first quenched product with ethyl acetate, thereby producing a first extracted product; and triturating the first extracted product with chloroform, thereby isolating the terpene trilactone GA in at least 98% purity and producing a filtrate;

15 d) concentrating the filtrate of step c) under vacuum to produce a concentrated filtrate; subjecting the concentrated filtrate to column chromatography with hexane/ethyl acetate, thereby separating benzylated terpene trilactone GB from residual terpene trilactone GA; deprotecting the separated terpene trilactone GB by catalytic hydrogenation, thereby obtaining terpene trilactone GB in at least 98% purity;

20 e) removing solvent under vacuum from the third fraction of step a) so as to produce a second

residue containing at least a preponderance of terpene trilactones GJ and GC; admixing the second residue in DMF with K_2CO_3 and benzyl bromide, thereby producing a second reaction mixture; quenching the second reaction mixture with HCl, thereby producing a second quenched product; extracting the second quenched product with ethyl acetate, thereby producing a second extracted product; subjecting the second extracted product to column chromatography with hexane/ethyl acetate, thereby separating benzylated terpene trilactone GC from terpene trilactone GJ, providing terpene trilactone GJ in at least 85% purity; recrystallizing terpene trilactone GJ with ethanol/water, thereby providing terpene trilactone GJ in at least 98% purity; and

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f) deprotecting the benzylated terpene trilactone GC of step e) by catalytic hydrogenation in ethanol, thereby obtaining terpene trilactone GC in at least 95% purity.

The invention also provides a process for obtaining terpene trilactones from Ginkgo biloba plant material or an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

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a) extracting the Ginkgo biloba plant material or the Ginkgo biloba extract with a first suitable solvent to produce a first residue and a first filtrate; extracting the first filtrate with a second suitable solvent to produce a second residue and a second filtrate;

b) subjecting the second filtrate to column chromatography with a first chromatography system to obtain Bilibalide;

- c) extracting the second residue with a third suitable solvent to obtain terpene trilactone Ginkgolide B (GB) and a third filtrate;
- d) subjecting the third filtrate to column chromatography with a second chromatography system to produce a first fraction containing terpene trilactone Ginkgolide A (GA) and GB and a second fraction containing terpene trilactone Ginkgolide C (GC) and terpene trilactone Ginkgolide J (GJ);
- e) subjecting the first fraction to iterative extractions with a fourth suitable solvent to separate GA and GB; and
- f) subjecting the second fraction to column chromatography with a third chromatography system to separate GC and GJ,
thereby obtaining terpene trilactones, Bilibalide, GA, GB, GC and GJ isolated from each other and from the Ginkgo biloba plant material or the extract of Ginkgo biloba.

The invention also provides a process for obtaining terpene trilactones from Ginkgo biloba plant material or an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

- a) extracting the Ginkgo biloba plant material or the Ginkgo biloba extract with ethyl acetate to produce a first residue and a first filtrate;
- b) concentrating the first filtrate of step a) to produce a concentrated first filtrate; extracting the concentrated first filtrate with diethyl ether to produce a second residue and a second filtrate;

5 c) concentrating the second filtrate of step b) to produce a second concentrated filtrate; subjecting the second concentrated filtrate to column chromatography with hexanes/ethyl acetate to produce a fraction containing the terpene trilactone Bilibalide;

10 d) removing the solvent from the fraction of step c) thereby obtaining the terpene trilactone Bilibalide;

15 e) admixing the second residue of step b) with methanol and filtering to produce a third filtrate and the terpene trilactone Ginkgolide B (GB);

20 f) concentrating the third filtrate of step e) to produce a third concentrated filtrate; subjecting the third concentrated filtrate to column chromatography with hexanes/acetone to produce a first fraction containing a mixture of terpene trilactones Ginkgolide A (GA) and GB and a second fraction containing a mixture of terpene trilactones Ginkgolide C (GC) and Ginkgolide J (GJ);

25 g) removing the solvent from the second fraction of step f) to produce a third residue; subjecting the third residue to column chromatography with diethyl ether/methanol to produce third fraction containing GC and a fourth fraction containing GJ;

30 h) removing the solvent from the third fraction of step g) to thereby obtain GC;

 i) removing the solvent from the fourth fraction of step b) to thereby obtain GJ;

 j) removing the solvent from the first fraction of step f) to produce a fourth residue; extracting

the fourth residue with methanol to produce GB and a fourth filtrate;

5 k) concentrating the fourth filtrate of step j) to produce a fourth concentrated filtrate; admixing the fourth concentrated filtrate with methanol and filtering to produce a fifth residue, which residue is GB enriched and a fifth filtrate;

10 l) concentrating the fifth filtrate of step k) and to produce a fifth concentrated filtrate; admixing the fifth concentrated filtrate with methanol and filtering to produce an sixth residue, which is GA enriched and a sixth filtrate;

15 m) concentrating the sixth filtrate of step l) to produce a sixth concentrated filtrate; admixing the sixth concentrated filtrate with methanol and crystallizing to produce GA;

20 n) admixing the fifth residue of step k) and the sixth residue of step l) to produce a first residue mixture, which is enriched in GA and GB; admixing the first residue mixture with methanol and filtering to produce GB and a seventh filtrate;

25 o) concentrating the seventh filtrate of step m) to produce a seventh concentrated filtrate; admixing the seventh concentrated filtrate with methanol and filtering to produce an eighth residue, which contains GA and an eighth filtrate;

30 p) concentrating the eighth filtrate of step o) to produce a eighth concentrated filtrate; admixing the eighth concentrated filtrate with methonal and crystallizing to produce GA,

- q) collecting the GA from steps m), o), and p), thereby obtaining GA; and
- r) collecting the GB from steps e), j), and n), thereby obtaining GB,

5 thereby obtaining terpene trilactones.

The invention also provides a process for separating each of Bilibalide, Ginkgolide A, (GA), Ginkgolide B, (GB), Ginkgolide C, (GC), and Ginkgolide J, (GJ) from a mixture of terpene 10 trilactones (TTL), wherein the separation is achieved through non-covalent interaction with the mixture of TTLs.

The invention also provides a process for isolating bilibalide from a mixture of terpene trilactones (TTL) wherein the 15 mixture comprises Bilibalide, Ginkgolide A, (GA), Ginkgolide B, (GB), Ginkgolide C, (GC), and Ginkgolide J, (GJ), the process comprising the steps of:

- a) extracting the mixture of TTLs with diethyl ether solvent to produce a residue and a filtrate; and
- b) subjecting the filtrate to column chromatography with a hexanes/ethyl acetate solvent system to thereby isolate Bilibalide 20 from the mixture of TTLs.

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Detailed Description of the Figures

Figure 1. Bilobalide and Ginkgolides A, B, C, and J.

5 **Figure 2.** Flow diagram showing the separation and isolation of
terpene trilactones from enriched *Ginkgo biloba* extracts by a
first method.

10 **Figure 3.** Evaluation of different alkyl groups for selective
alkylation of ginkgolides B and C.

Figure 4. Hydrogenolysis of benzylated ginkgolides B and C.

15 **Figure 5.** Flow diagram showing the speration and isolation of
terpene trilactones from Bioginkgo® 27/7 extract by a second
method.

Detailed Description of the Invention

The subject invention provides a process for separating a terpene trilactone from Ginkgo biloba plant material or from
5 an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

10 a) subjecting the Ginkgo biloba plant material or the extract to column chromatography with an appropriate solvent system to produce at least a first fraction containing the terpene trilactone bilobalide, a second fraction eluted after the first fraction containing the terpene trilactones GA and GB, and a third fraction eluted after the second fraction containing at least a preponderance of the terpene trilactones GC and GJ; and

15 b) alkylating the terpene trilactone GB of the second fraction so as to produce a first mixture including terpene trilactone GA and alkylated terpene trilactone GB; or alkylating the terpene trilactone GC of the third fraction so as produce a second mixture including terpene trilactone GJ and alkylated terpene trilactone GC, so as to thereby
20 isolate a terpene trilactone.

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In one embodiment, in step a) of the process, the solvent system is hexane/ethyl acetate or petroleum ethers/ethyl acetate.

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In another embodiment the process further comprises triturating the first fraction with a suitable solvent to produce the terpene trilactone bilobalide in at least 95% purity.

In yet another embodiment, the terpene trilactone bilobalide is produced in at least 98% purity.

5 In yet another embodiment, the suitable solvent is ethyl ether.

In a further embodiment, the alkylation of the terpene trilactone GB of the second fraction comprises the steps of:

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a) removing solvent from the second fraction so as to produce a first residue containing at least a preponderance of terpene trilactones GA and GB;

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b) admixing the first residue in a suitable polar solvent with a suitable base and R-Br,

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where R is a substituted or unsubstituted, straight-branched, or cyclic alkyl, or $\text{-CH}_{(2-5)}\text{-Ar}$,

where Ar is an aromatic group,

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so as to produce a first reaction mixture;

c)

quenching the first reaction mixture of step b) with a suitable acid;

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d) extracting the product of step c) with a first suitable solvent so as to produce a composition comprising terpene trilactone GA and alkylated terpene trilactone GB; and

e) triturating the product of step d) with a second suitable solvent, thereby isolating the terpene trilactone GA in at least 95% purity and producing a filtrate containing alkylated terpene trilactone GB.

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In yet a further embodiment, in step e) of the process, the terpene trilactone GA is isolated in at least 98% purity.

In still yet a further embodiment of the process wherein the 10 terpene trilactone GB of the second fraction is alkylated,

in step a), the solvent is removed under vacuum;

15 in step b), the suitable polar solvent is DMF, the suitable base is K_2CO_3 or Cs_2CO_3 , and R-Br is benzyl bromide;

in step c), the suitable acid is HCl;

20 in step d), the first suitable solvent is ethyl acetate; and

in step e), the second suitable solvent is chloroform.

25 In another embodiment the process further comprises the steps of:

f) concentrating the filtrate of step e) under vacuum to produce a concentrated filtrate;

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g) subjecting the concentrated filtrate to column chromatography with a suitable solvent system, thereby separating alkylated terpene trilactone GB from residual terpene trilactone GA; and

5 h) deprotecting the alkylated terpene trilactone GB of step g) by catalytic hydrogenation, thereby obtaining the terpene trilactone GB in at least 95% purity.

In yet another embodiment, in step h) the terpene trilactone GB is obtained in least 98% purity.

10 10 In still yet another embodiment, the suitable solvent system in step g) is hexane/ethyl acetate or petroleum ethers/ethyl acetate.

15 15 In even yet another embodiment, the alkylation of the terpene trilactone GC of the third fraction comprises the steps of:

20 a) removing solvent from the third fraction so as to produce a second residue containing at least a preponderance of terpene trilactones GJ and GC;

b) admixing the second residue in a suitable polar solvent with a suitable base and R-Br,

25 25 where R is a substituted or unsubstitued, straight-chained or branched, or cyclic alkyl, or $-\text{CH}_{(2-5)}-\text{Ar}$,

where Ar is an aromatic group,

30 so as to produce a second reaction mixture;

c) quenching the second reaction mixture of step b) with a suitable acid;

d) extracting the product of step c) with a suitable solvent so as to produce a composition comprising terpene trilactone GJ and alkylated terpene trilactone GC;

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e) subjecting the product of step d) to column chromatography with a first suitable solvent system, thereby separating alkylated terpene trilactone GC from terpene trilactone GJ, and providing terpene trilactone GJ in at least 85% purity; and

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f) recrystallizing terpene trilactone GJ from step e) with a second suitable solvent system, thereby providing terpene trilactone GJ in at least 95% purity.

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In another embodiment of the alkylation of the terpene trilactone GC of the third fraction,

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in step a), the solvent is removed under vacuum;

in step b), the suitable polar solvent is DMF, the suitable base is K_2CO_3 or Cs_2CO_3 , and R-Br is benzyl bromide;

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in step c), the acid is HCl;

in step d), the solvent is ethyl acetate;

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in step e), the first suitable solvent system is hexane/ethyl acetate or petroleum ethers/ethyl acetate; and

in step f), the second suitable solvent system is ethanol/water.

In even yet another embodiment, the process comprises the step
5 of deprotecting the alkylated terpene trilactone GC of the third fraction by catalytic hydrogenation in a suitable solvent, thereby obtaining the terpene trilactone GC in at least 95% purity.

10 In still yet another embodiment, the terpene trilactone GC is obtained in at least 98% purity.

In a further embodiment, the suitable solvent in the process for deprotecting the alkylated terpene trilactone GC of the
15 third fraction is ethanol.

The invention further provides a composition comprising terpene trilactones GA and GB of step a) of the described process.

20 In another embodiment, the invention provides a composition comprising terpene trilactone GA and alkylated terpene trilactone GB of step d) of the process wherein terpene trilactone GB of the second fraction is alkylated.

25 In yet another embodiment, the invention yet further provides a composition comprising terpene trilactones GC and GJ of step a) of the process wherein terpene trilactone GC of the third fraction is alkylated.

30 In still yet another embodiment, invention even further provides a composition comprising alkylated terpene trilactone GC and terpene trilactone GJ present of step d) of of the

process wherein terpene trilactone GC of the third fraction is alkylated.

The invention also provides a product prepared by the process
5 of step a) of the process wherein terpene trilactone GB of the second fraction is alkylated.

In a further embodiment, the invention provides a product prepared by step d) of the process wherein terpene trilactone
10 GB of the second fraction is alkylated.

In still a further embodiment, the invention provides a product prepared by step a) of the process wherein terpene trilactone GC of the third fraction is alkylated.

15 In yet a further embodiment, the invention provides a product prepared by the step d) of the process wherein terpene trilactone GC of the third fraction is alkylated.

20 The invention also provides a process for separating a mixture of terpene trilactones GA and GB comprising the steps of:

- a) dissolving the mixture in a suitable solvent;
- 25 b) adding benzyl bromide and a suitable base to the mixture, thereby preferentially benzylating terpene trilactone GB;
- c) separating benzylated terpene trilactone GB from terpene trilactone GA by column chromatography; and
- 30 d) deprotecting benzylated terpene trilactone GB by catalytic hydrogenation.

The invention also provides a process for separating a mixture of terpene trilactones GC and GJ comprising the steps of:

- a) dissolving the mixture in a suitable solvent;
- 5 b) adding benzyl bromide and a suitable base to the mixture, thereby preferentially benzylating terpene trilactone GC;
- 10 c) separating benzylated terpene trilactone GC from terpene trilactone GJ by column chromatography; and
- d) deprotecting benzylated terpene trilactone GC by catalytic hydrogenation.

15 The invention also provides a process for separating a terpene trilactone from Ginkgo biloba plant material or an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the process comprising the steps of:

- 20 a) subjecting the Ginkgo biloba plant material or the extract to column chromatography with hexane/ethyl acetate to produce at least a first fraction containing the terpene trilactone bilobalide, a second fraction eluted after the first fraction containing the terpene trilactone GA and GB, and a third fraction eluted after the second fraction containing at least a preponderance of the terpene trilactones GC and GJ;
- 25 b) rinsing the first fraction of step a) with ethyl ether to produce the terpene trilactone bilobalide in at least 98% purity;

5 c) removing solvent under vacuum from the second fraction in step a) so as to produce a first residue containing at least a preponderance of terpene trilactones GA and GB; admixing the first residue in DMF with K₂CO₃ and benzyl bromide, thereby producing a first reaction mixture; quenching the first reaction mixture with HCl, thereby producing a first quenched product; extracting the first quenched product with ethyl acetate, thereby producing a first extracted product; and triturating the first extracted product with chloroform, thereby isolating the terpene trilactone GA in at least 98% purity and producing a filtrate;

10 d) concentrating the filtrate of step c) under vacuum to produce a concentrated filtrate; subjecting the concentrated filtrate to column chromatography with hexane/ethyl acetate, thereby separating benzylated terpene trilactone GB from residual terpene trilactone GA; deprotecting the separated terpene trilactone GB by catalytic hydrogenation, thereby obtaining terpene trilactone GB in at least 98% purity;

15 e) removing solvent under vacuum from the third fraction of step a) so as to produce a second residue containing at least a preponderance of terpene trilactones GJ and GC; admixing the second residue in DMF with K₂CO₃ and benzyl bromide, thereby producing a second reaction mixture; quenching the second reaction mixture with HCl, thereby producing a second quenched product; extracting the second quenched product with ethyl acetate, thereby producing a second extracted product; subjecting the

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second extracted product to column chromatography with hexane/ethyl acetate, thereby separating benzylated terpene trilactone GC from terpene trilactone GJ, providing terpene trilactone GJ in at least 85% purity; recrystallizing terpene trilactone GJ with ethanol/water, thereby providing terpene trilactone GJ in at least 98% purity; and

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10 f) deprotecting the benzylated terpene trilactone GC of step e) by catalytic hydrogenation in ethanol, thereby obtaining terpene trilactone GC in at least 95% purity.

In yet another embodiment, the invention provides a process
15 wherein terpene trilactone GJ in step g) is provided in at least 90% purity.

In still another embodiment, the invention provides a process
wherein terpene trilactone GC in step f) is obtained in at
20 least 98% purity.

The invention also provides a process for obtaining terpene trilactones from Ginkgo biloba plant material or an extract of Ginkgo biloba comprising a mixture of terpene trilactones, the
25 process comprising the steps of:

30 a) extracting the Ginkgo biloba plant material or the Ginkgo biloba extract with a first suitable solvent to produce a first residue and a first filtrate; extracting the first filtrate with a second suitable solvent to produce a second residue and a second filtrate;

b) subjecting the second filtrate to column chromatography with a first chromatography system to obtain Bilibalide;

c) extracting the second residue with a third suitable solvent to obtain terpene trilactone Ginkgolide B (GB) and a third filtrate;

5 d) subjecting the third filtrate to column chromatography with a second chromatography system to produce a first fraction containing terpene trilactone Ginkgolide A (GA) and GB and a second fraction containing terpene trilactone Ginkgolide C (GC) and terpene trilactone Ginkgolide J (GJ);

10 e) subjecting the first fraction to iterative extractions with a fourth suitable solvent to separate GA and GB; and

15 f) subjecting the second fraction to column chromatography with a third chromatography system to separate GC and GJ,

thereby obtaining terpene trilactones, Bilibalide, GA, GB, GC and GJ isolated from each other and from the Ginkgo biloba plant material or the extract of Ginkgo
20 biloba.

In another embodiment, the first suitable solvent is ethyl acetate.

25 In yet another embodiment, the second suitable solvent is diethyl ether.

In still another embodiment, the third suitable solvent is methanol.

30 In another embodiment, the fourth suitable solvent is methanol.

In another embodiment, the first chromatography system comprises hexanes/ethyl acetate.

In another embodiment, the second chromatography system
5 comprises hexanes/acetone.

In yet another embodiment, the second chromatography system comprises diethyl ether/methanol.

10 In still another embodiment, subjecting the second filtrate to chromatography comprises the steps of:

- a) concentrating the second filtrate to produce a second concentrated filtrate;
- b) subjecting the second concentrated filtrate to column chromatography with hexanes/ethyl acetate to produce a fraction containing the terpene trilactone Bilibalide; and
- c) removing the solvent from the fraction of step b) thereby obtaining the terpene trilactone Bilibalide.

20 In another embodiment, subjecting the third filtrate to chromatography comprises the steps of:

- a) concentrating the third filtrate to produce a third concentrated filtrate; and
- b) subjecting the third concentrated filtrate to column chromatography with hexanes/acetone to produce a first fraction containing a mixture of terpene trilactones Ginkgolide A (GA) and GB and a second fraction containing a mixture of terpene trilactones Ginkgolide C (GC) and Ginkgolide J (GJ).

In yet another embodiment, extracting the fraction containing a mixture of GA and GB with a fourth suitable solvent to obtain GA and GB comprises the steps of:

- a) removing the solvent from the fraction to produce a residue;
- b) extracting the residue of step a) with methanol to produce a filtrate and GB;
- c) concentrating the filtrate of step b) to produce a concentrated filtrate of step b); admixing the concentrated filtrate of step b) with methanol and filtering to produce a GB enriched residue and a filtrate;
- d) concentrating the filtrate of step c) to produce a concentrated filtrate of step c); admixing the concentrated filtrate of step c) with methanol and filtering to produce a GA enriched residue and a filtrate;
- e) concentrating the filtrate of step d) to produce a concentrated filtrate of step d); admixing the concentrated filtrate of step d) with methanol and crystallizing to produce GA;
- f) admixing the GB enriched residue of step c) and the GA residue of step d) to produce a first residue mixture, which is enriched in GA and GB; admixing the first residue mixture with methanol and filtering to produce GB and a filtrate;
- g) concentrating the filtrate of step f) to produce a concentrated filtrate of step f); admixing the concentrated filtrate of step f) with methanol and filtering to produce GA and a filtrate;
- h) concentrating the filtrate of step g) to produce a concentrated filtrate of step g);

admixing the concentrated filtrate of step g) with methanol and crystallizing to produce GA,

- i) collecting GA from steps e), g), and h), thereby obtaining GA; and
- j) collecting the terpene trilacton GB from steps b), and f), thereby obtaining GB.

5 In another embodiment, subjecting the fraction containing GC and GJ to chromatography comprises the steps of:

- a) concentrating the fraction to produce a concentrated fraction;
- b) subjecting the concentrated fraction to column chromatography with diethyl ether/methanol to produce a GC containing fraction and a GJ containing fraction;
- c) removing the solvent from the GC containing fraction thereby obtaining GC; and
- 10 d) removing the solvent from the GJ containing fraction thereby obtaining GJ.

15 The invention also provides a process for obtaining terpene trilactones from *Ginkgo biloba* plant material or an extract of *Ginkgo biloba* comprising a mixture of terpene trilactones, the process comprising the steps of:

- a) extracting the *Ginkgo biloba* plant material or the *Ginkgo biloba* extract with ethyl acetate to produce a first residue and a first filtrate;
- 20 b) concentrating the first filtrate of step a) to produce a concentrated first filtrate; extracting the concentrated first filtrate with diethyl ether to produce a second residue and a second filtrate;
- c) concentrating the second filtrate of step b) to produce a second concentrated filtrate;

subjecting the second concentrated filtrate to column chromatography with hexanes/ethyl acetate to produce a fraction containing the terpene trilactone Bilibalide;

5 d) removing the solvent from the fraction of step c) thereby obtaining the terpene trilactone Bilibalide;

10 e) admixing the second residue of step b) with methanol and filtering to produce a third filtrate and the terpene trilactone Ginkgolide B (GB);

15 f) concentrating the third filtrate of step e) to produce a third concentrated filtrate; subjecting the third concentrated filtrate to column chromatography with hexanes/acetone to produce a first fraction containing a mixture of terpene trilactones Ginkgolide A (GA) and GB and a second fraction containing a mixture of terpene trilactones Ginkgolide C (GC) and Ginkgolide J (GJ);

20 g) removing the solvent from the second fraction of step f) to produce a third residue; subjecting the third residue to column chromatography with diethyl ether/methanol to produce third fraction containing GC and a fourth fraction containing GJ;

25 h) removing the solvent from the third fraction of step g) to thereby obtain GC;

30 i) removing the solvent from the fourth fraction of step b) to thereby obtain GJ;

 j) removing the solvent from the first fraction of step f) to produce a fourth residue; extracting the fourth residue with methanol to produce GB and a fourth filtrate;

5 k) concentrating the fourth filtrate of step j) to produce a fourth concentrated filtrate; admixing the fourth concentrated filtrate with methanol and filtering to produce a fifth residue, which residue is GB enriched and a fifth filtrate;

10 l) concentrating the fifth filtrate of step k) and to produce a fifth concentrated filtrate; admixing the fifth concentrated filtrate with methanol and filtering to produce an sixth residue, which is GA enriched and a sixth filtrate;

15 m) concentrating the sixth filtrate of step l) to produce a sixth concentrated filtrate; admixing the sixth concentrated filtrate with methanol and crystallizing to produce GA;

20 n) admixing the fifth residue of step k) and the sixth residue of step l) to produce a first residue mixture, which is enriched in GA and GB; admixing the first residue mixture with methanol and filtering to produce GB and a seventh filtrate;

25 o) concentrating the seventh filtrate of step m) to produce a seventh concentrated filtrate; admixing the seventh concentrated filtrate with methanol and filtering to produce an eighth residue, which contains GA and an eighth filtrate;

30 p) concentrating the eighth filtrate of step o) to produce a eighth concentrated filtrate; admixing the eighth concentrated filtrate with methonal and crystallizing to produce GA,

 q) collecting the GA from steps m), o), and p), thereby obtaining GA; and

r) collecting the GB from steps e), j), and n),
thereby obtaining GB,
thereby obtaining terpene trilactones.

5 The invention also provides a process for separating each of Bilibalide, Ginkgolide A, (GA), Ginkgolide B, (GB), Ginkgolide C, (GC), and Ginkgolide J, (GJ) from a mixture of terpene trilactones (TTL), wherein the separation is achieved through non-covalent interaction with the mixture of TTLs.

10

The invention also provides a process for isolating bilibalide from a mixture of terpene trilactones (TTL) wherein the mixture comprises Bilibalide, Ginkgolide A, (GA), Ginkgolide B, (GB), Ginkgolide C, (GC), and Ginkgolide J, (GJ), the process comprising the steps of:

15

- a) extracting the mixture of TTLs with diethyl ether solvent to produce a residue and a filtrate; and
- b) subjecting the filtrate to column chromatography with a hexanes/ethyl acetate solvent system to thereby isolate Bilibalide from the mixture of TTLs.

20
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Experimental Details

In the following examples, the invention is described in detail. The examples describe a method for isolating five 5 terpene trilactones from *G. biloba*. The terpene trilactones were each isolated in high purity. Besides providing each terpene trilactone in higher purity and higher yield, the disclosed method involves a minimum of steps.

10 General Experimental Procedures

Reactions were monitored by analytical TLC with silica gel 60 F₂₅₄ and spots were visualized by heating and 254 nm irradiation. Column chromatography was performed using silica gel (230-400 mesh). ¹H-NMR spectra were recorded on Bruker 15 (300, 400 or 500 MHz) spectrometers. Purity was estimated from ¹H-NMR spectra. Mass spectra were measured on JEOL JMS-HX110/100A HF mass spectrometer under FAB conditions with NBA as the matrix.

20 Extract Material

Enriched extract was prepared according to the method disclosed in U.S. Patent Application No. 6,590,109 starting with Bioginkgo 27/7 extract (Pharmanex, Provo, UT). All other reagents were purchased and used as received.

25

EXAMPLE 1

Chromatography of Enriched *G. biloba* Extract. The enriched extract (4.0 g) in minimum amount of EtOAc was loaded on a 30 silica gel (100 g) column. The column was slowly eluted with EtOAc/hexanes solvent mixtures either with or without slight external pressure. The initial solvent system was EtOAc/hexanes (3.5:6.5, 500 mL). Content of EtOAc in the eluent was increased gradually in six steps (4:6, 500 mL;

4.5:5.5, 500 mL; 5:5, 500 mL; 5.5:4.5, 500 mL; 6:4, 400 mL; 6.5:3.5, 400 mL). Content of the collected fractions was analyzed by TLC, and if necessary, was also analyzed by ¹H-NMR. The fractions collected at 45% EtOAc/hexanes contained 5 bilobalide (0.4 g). The fractions collected at 50% EtOAc/hexanes contained small amounts of impure BB and GA then mixture GA/GB. The fractions collected at 55% EtOAc/hexanes contained mixture GA/GB (1.1 g). The fractions collected at 60% EtOAc/hexanes contained mixture of GC/GJ (0.4 g) with 10 small amount of GA and GB.

EXAMPLE 2

Purification of bilobalide. Bilobalide (310 mg) from the chromatography above was suspended in 2 mL of ethyl ether, 15 filtered and washed twice with 1 mL of ethyl ether to yield 217 mg of bilobalide as a white powder (purity by ¹H-NMR \geq 98%).

EXAMPLE 3

Benzylation of a mixture GA/GB. To a ginkgolide mixture from the above chromatography, (1.08 g, GB ~25% w/w, 0.63 mmol of GB; GA ~74% w/w) was added K₂CO₃ (879 mg, 6.3 mmol, 10 eq.) and DMF (11 mL). While stirring, benzyl bromide (756 mL, 6.3 mmol, 10 eq.) was added. The mixture was stirred for 40 minutes, then quenched with 1M HCl (18 mL) and the solution 25 was extracted with EtOAc (3 x) and dried with MgSO₄. Volatiles were removed under reduced pressure and/or by multiple azeotropic evaporations with chloroform. The product mixture was suspended in chloroform (10mL), filtered and washed twice with chloroform (4mL and 2 mL) to obtain 605 mg of GA (2) as a 30 white powder (purity by ¹H-NMR \geq 98%). The filtrate was concentrated and purified by gradient column chromatography (30 - 50% EtOAc/hexanes) to obtain 326 mg of 6 and 134 mg of GA (2). GA from the chromatography was further purified by recrystallization (EtOH/H₂O). All analytical data as previously

reported (Roumestand, C.; Perly, B.; Hosford, D.J.; Braquet, P. *Tetrahedron* **1989**, *45*, 1975-1989).

EXAMPLE 4

5 **Benzylation of a mixture GA/GB/GC/GJ.** To a ginkgolide mixture (543mg, GA~3%, GB~4.5%, GC~68%, GJ~21% w/w, 0.92mmol of GB+GC) was added K₂CO₃ (1.27g, 9.2mmol, 10eq.) and DMF (18mL). While stirring, benzyl bromide (1.09mL, 9.2mmol, 10eq.) was added. The mixture was stirred for 2 hours then quenched with 1M HCl (30mL) and the solution was extracted with EtOAc (3 x) and dried with MgSO₄. Volatiles were removed under reduced pressure and by multiple azeotropic evaporations with chloroform. The mixture was purified by gradient column chromatography (30 - 60% EtOAc/hexanes) to obtain 31.2 mg of **6**, 407.7 mg of **7**, 18.6 mg of GA (**2**) and 111 mg of GJ (**5**). GA and GJ were further purified by recrystallization from EtOH/H₂O. All analytical data as previously reported (Roumestand, C.; Perly, B.; Hosford, D.J.; Braquet, P. *Tetrahedron* **1989**, *45*, 1975-1989; Hu, L.; Chen, Z.; Xie, Y.; Jiang, H.; Zhen, H. *Bioorg. Med. Chem.* **2000**, *8*, 1515-1521; Vogensen, S.B.; Stromgaard, K.; Shindou, H.; Jaracz, S.; Suehiro, M.; Ishii, S.; Shimizu, T.; Nakanishi, K. *Journal of Medicinal Chemistry* **2003**, *46*, 601-608).

25 **EXAMPLE 5**

Hydrogenation of 6. To a solution of **6** (322 mg, 0.626 mmol) in EtOH (5 mL) in a thick-wall flask was added 10% Pd/C (64 mg) and the mixture was vigorously stirred under H₂ (56 psi, 3.8 atm). After 24 hours, the reaction mixture was filtered through celite. Volatiles were removed under reduced pressure to obtain 257 mg of ginkgolide B as a slightly brownish crystalline powder (97% yield). All analytical data as previously reported (purity by ¹H-NMR >98%) (Roumestand, C.;

Perly, B.; Hosford, D.J.; Braquet, P. *Tetrahedron* 1989, 45, 1975-1989).

EXAMPLE 6

5 **Hydrogenation of 7:** To a solution of 7 (153 mg, 0.288 mmol) in EtOH (4 mL) in a thick-wall flask was added 10% Pd/C (31 mg) and the mixture was vigorously stirred under H₂ (56 psi, 3.8 atm). After 20 hours, the reaction mixture was filtered through celite. Volatiles were removed under reduced pressure.

10 The product was washed with 15% EtOAc/hexanes (2 x 5 mL) to obtain 121.7 mg (96% yield) of GC (4) as a white powder. All analytical data as previously reported (purity by ¹H-NMR >98%) (Roumestand, C.; Perly, B.; Hosford, D.J.; Braquet, P. *Tetrahedron* 1989, 45, 1975-1989).

15

RESULTS

An efficient and rapid protocol has been developed for separation of individual ginkgolides and bilobalide from *Ginkgo Biloba* extracts. The procedure takes advantage of the enhanced susceptibility for benzylation that exists in ginkgolides B and C, contrasted with the unreactivity of ginkgolides A and J. The protocol is applicable to enriched extracts prepared from *Ginkgo Biloba* leaves as described previously (U.S. Patent No. 6,590,109, issued July 8, 2003 to Lichtblau, et al.). Bilobalide is separated by a single chromatography prior to benzylation. After derivatization, the ginkgolides are separated by a single column chromatography, and ginkgolides B and C are obtained from their benzylated derivatives by hydrogenolysis.

30

In 1992, Corey converted ginkgolides B and C which were in a mixture with ginkgolide A to benzyloxymethyl ethers, separated them by column chromatography and removed the protecting group by hydrogenolysis (Corey, E.J.; Rao, K.S.; Ghosh, A. K.

Tetrahedron Lett. 1992, 33, 6955-6958). Only ginkgolides B and C undergo etherification because of the presence of 1-OH, which stabilizes the alkoxy anion from 10-OH by hydrogen bonding. The method described herein is a gram-scale protocol in which the individual ginkgolides A/B/C/J and bilobalide are isolated and purified rapidly and efficiently from a crude G. biloba extract by a combination of benzylation, chromatography, and hydrogenolysis (Figure 2).

10 Initially, the enriched extract (Lichtblau, D., Berger, J.M.; Nakanishi, K. Journal of Natural Products 2002, 65, 1501-1504) (53% TTLs) was subjected to a column chromatography on silica gel, with EtOAc/hexanes as a solvent system for elution (gradient, 35 - 65% EtOAc/hexanes) either with or without slight external pressure. This chromatography is necessary because bilobalide is unstable in conditions used during alkylation. Bilobalide thus obtained was already ~90% pure. Trituration with ethyl ether provided pure bilobalide (>98%) as a white powder. Surprisingly, the single column chromatography also yielded a nearly baseline separation of ginkgolide pairs GA/GB and GC/GJ with dramatically reduced content of flavonoids and other components of the extract. The later pair contained also small amounts of GA and GB.

25 Several alkyl groups for selective alkylation of GB/GC in the presence of GA/GJ were studied (Figure 3). Interestingly, reaction with allyl bromide or cinnamyl bromide lead to partial alkylation of GA (2). However, the 10-alkyl-GA derivatives could not be separated from the 10-alkyl-GB derivatives. Benzyl bromide reacted with GA very slowly and no product could be detected with the first 90 minutes. Therefore, benzylation was selected for further consideration since GA/GJ do not react, while an excellent separation of benzyl ginkgolides 10-Bn-GB, 6 and 10-Bn-GC, 7 from unreacted

GA/GJ is achieved on silica gel, and the benzyl groups can be readily removed by hydrogenolysis on Pd/C.

Of the bases we tested, the best results were obtained by
5 using K_2CO_3 and Cs_2CO_3 as base; Na_2CO_3 and Et_3N lead to incomplete conversion while stronger bases such as alkali hydrides or LDA lead to decomposition and a complex mixture of products. Of the solvents we tested, DMF was the best solvent because of quantitative conversions and no side reactions.
10 The benzylation in other polar solvents such as acetone, acetonitrile and methanol lead to lower yields forming various side products, which could not be removed by chromatography. Increase in temperature or reaction time to more than 3 hours lead to formation of 10-Bn-GA (8), which is inseparable from
15 10-Bn-GB (6). Since benzyl derivatives are soluble in halogenated hydrocarbons, mixture of 10-Bn-GB (6) and GA (2) could be separated by trituration with chloroform. Ginkgolide A thus obtained as a white powder had purity of >98%. The filtrate, rich in 10-Bn-GB (6), was further purified by column
20 chromatography on silica gel to obtain pure 10-BnGB (6) and GA (2). The mixture after benzylation of GC/GJ (with traces of GA/GB) was separated directly by column chromatography to obtain 10-Bn-GB (6), 10-Bn-GC (7), GA (2) and GJ (5). Catalytic hydrogenation (10%Pd/C) of the benzyl ethers under 4
25 atm of H_2 effected quantitative deprotection in 24 h to obtain native GB (3) and GC (4) (Figure 4). Interestingly, 10-Bn-GA (8) as the impurity in some samples of 10-Bn-GB (6), did not react under these conditions, therefore GB (3) was not contaminated by GA (2) and the purification was facile. It is
30 likely that 1-OH in 10-Bn-GB (6) and 10-Bn-GC (7) coordinates to Pd during hydrogenation. This activation is not possible in the case of GA derivative (8) therefore Pd cannot reach the sterically hindered benzyl group. To obtain pure compounds,

the ginkgolides were further recrystallized in some cases (EtOH/H₂O).

In conclusion, we have developed a simple and efficient method
5 for isolation of five TTLs from *G. biloba* extracts on gram scale. Starting from enriched extracts with >50% of TTLs, only three column chromatographies suffices. Benzylation of the ginkgolides can be performed in reagent grade DMF on air. Trituration affords GA in excellent purity. All of this is
10 possible without resorting to expensive preparative reversed-phase HPLC. This is of particular importance for large-scale isolation of ginkgolides as phytopharmaceuticals. Furthermore, 10-Bn-GC (7) is an ideal precursor for a wide variety of transformations including the transformation into
15 GB(Corey, E.J.; Rao, K.S.; Ghosh, A. K. *Tetrahedron Lett.* 1992, 33, 6955-6958; Teng, B.P. in *Ger. Offen.*; (Societe De Conseils De Recherches Et D'applications Scientifiques (SCRAS), Fr.). De, 1992, p 6).

EXAMPLE 7

Isolation of Ginkgolides from Bioginkgo® 27/7 extract. As shown in Figure 5, Bioginkgo® extract 27/7(100 g) was extracted with EtOAc (800 ml). The EtOAc extract was evaporated to give 7.0 g of brown solid. This solid was washed with Et₂O (200 ml). The Et₂O extract was concentrated and purified by column chromatography (SiO₂, 100g, hexanes-EtOAc, 2:1) to give Bilibalide (0.7 g). The residue was dissolved in MeOH (20 ml) and filtrated to give colorless powder of Ginkgolide B (1.2 g). The filtrate was concentrated and subjected to silica gel column chromatography (SiO₂, 500 g, hexanes-acetone, 2:1) to give fractions of GA/GB mixture (2.3 g) and GC/GJ mixture (0.5 g). The GC/GJ mixture was purified by column chromatography (SiO₂, 200g, wet Et₂O-MeOH, 50:1) to give GC (0.4 g) and GJ (0.1 g). The GA/GB mixture was dissolved in MeOH and filtered to give GB (0.2 g) as a residue. The filtrate was concentrated and dissolved in MeOH, then filtered to give GB enriched residue. The filtrate was concentrated and dissolved in MeOH, then filtered to give GA enriched residue. The filtrate was crystallized with MeOH to give GA (1.3 g). The enriched GA and GB residues were combined and further purified by repeating the series of methanol extractions, filtrations and crystallizations to give GA (1.2 g) and GB (0.1 g), respectively.

25

RESULTS

This isolation of ginkgolides has been carried out using simple extraction, crystallization and silica gel column chromatography without chemical modification of ginkgolides or 30 using of reverse phase column chromatography.